

University of Dundee

Mammalian mitophagy – from in vitro molecules to in vivo models

Rodger, Catherine E.; McWilliams, Thomas G.; Ganley, Ian G.

Published in:
FEBS Journal

DOI:
[10.1111/febs.14336](https://doi.org/10.1111/febs.14336)

Publication date:
2018

Licence:
CC BY

Document Version
Publisher's PDF, also known as Version of record

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Rodger, C. E., McWilliams, T. G., & Ganley, I. G. (2018). Mammalian mitophagy – from *in vitro* molecules to *in vivo* models. *FEBS Journal*, 285(7), 1185-1202. <https://doi.org/10.1111/febs.14336>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Mammalian mitophagy – from *in vitro* molecules to *in vivo* models

Catherine E. Rodger, Thomas G. McWilliams and Ian G. Ganley

MRC Protein Phosphorylation and Ubiquitylation Unit, School of Life Sciences, University of Dundee, UK

Keywords

autophagy; development; disease; metabolism; mitochondria; mitophagy; *mito*-QC; mouse models; NIX; Parkin

Correspondence

I. G. Ganley, MRC Protein Phosphorylation and Ubiquitylation Unit, School of Life Sciences, University of Dundee, Dundee, DD1 5EH, UK
Fax: +44 1382 223778
Tel: +44 1382 388905
E-mail: i.ganley@dundee.ac.uk

The autophagic turnover of mitochondria, termed mitophagy, is thought to play an essential role in not only maintaining the health of the mitochondrial network but also that of the cell and organism as a whole. We have come a long way in identifying the molecular components required for mitophagy through extensive *in vitro* work and cell line characterisation, yet the physiological significance and context of these pathways remain largely unexplored. This is highlighted by the recent development of new mouse models that have revealed a striking level of variation in mitophagy, even under normal conditions. Here, we focus on programmed mitophagy and summarise our current understanding of why, how and where this takes place in mammals.

(Received 14 August 2017, revised 9 October 2017, accepted 14 November 2017)

doi:10.1111/febs.14336

Introduction

Mitochondrial homeostasis underpins cell survival. These multifunctional organelles are not only at the heart of cellular energy metabolism but also play key roles in many other processes, including calcium storage, iron metabolism and apoptosis. Maintaining a healthy mitochondrial population presents a huge cellular challenge and, as the primary sources of potentially hazardous reactive oxygen species and instigators of programmed cell death, failure to do so could have disastrous repercussions. This is especially important for the viability of long-lived, postmitotic cells such as neurons and cardiomyocytes, where the preservation

of mitochondria may underpin a healthy human lifespan. The accumulation of dysfunctional mitochondria has been tied to ageing and the progression of many human diseases, ranging from cancer to neurodegeneration. While cell signalling systems such as chaperones and proteases act to survey mitochondrial integrity, mitochondrial quality control also extends to the lysosome-dependent autophagic turnover of entire organelles. During the selective destruction of damaged mitochondria, termed mitophagy, a portion of the mitochondrial network is gradually engulfed within a double-membraned autophagosome, known as a

Abbreviations

AMPK, AMP-activated protein kinase; CL, cardiolipin; DCT, distal convoluted tubule; Drp1, dynamin-related protein 1; ER, endoplasmic reticulum; FUNDC1, FUN-14 domain containing protein 1; IMM, inner mitochondrial membrane; iPSC, induced pluripotent stem cell; KO, knockout; LIR, LC3-interacting region; MAM, mitochondria-associated ER membrane; mTORC1, mammalian target of rapamycin complex 1; OMM, outer mitochondrial membrane; OXPHOS, oxidative phosphorylation; PCT, proximal convoluted tubule; PHB2, prohibitin 2; PINK1, PTEN-induced putative kinase 1; PKA, protein kinase A; RGC, retinal ganglion cell; ULK1, UNC-51 like kinase 1.

mitophagosome. This eventually fuses with a compartment of the endolysosomal system, resulting in the proteolytic degradation of its mitochondrial cargo (Fig. 1). Notably, mitophagy is not the only lysosomal pathway implicated in mitochondrial turnover; for example, mitochondrial-derived vesicles (MDVs) can also deliver damaged mitochondrial components to the lysosome and are thus also implicated in quality control [1]. In this review, we will first discuss insights from cell-based studies that have provoked great interest in the significance of mitophagy as a protective pathway during health and disease. We then describe our current mechanistic understanding of how it is coordinated. Finally, we discuss the advent of new *in vivo* mitophagy models that reveal the striking instances of mitophagy that occur under steady-state physiological conditions.

Reasons to trigger mitophagy

Multiple mitophagy pathways operate within the cell and fulfil various roles. Mitophagy is thought to perform a housekeeping role under steady-state conditions, being responsible for the timely turnover of aged organelles. In addition, to combat the potential risks associated with dysfunctional mitochondria, mitophagy can also be rapidly enhanced as a protective mechanism to selectively remove damaged mitochondria and this has been extensively reviewed [2–4]. More recently, it has become apparent that commitment of

mitochondria to mitophagy-dependent degradation is a core mechanism by which organelle quantity and/or functional characteristics may be adjusted. This is a variant of mitophagy, referred to as programmed mitophagy [5], described to occur independently of mitochondrial damage and is engaged in an array of developmental and physiological contexts.

Important insights into programmed mitophagy were first recorded by the Ney laboratory while studying erythrocyte differentiation and maturation. Erythrocytes rapidly develop from immature erythroblasts generated in the bone marrow over a period of 48–72 h [6]. Erythroblasts are enucleated to form erythrocytes, before all remaining internal organelles and ribosomes are destroyed [6]. During this process, the specific removal of mitochondria occurs via mitophagy [7,8].

In addition to the role of mitophagy in erythrocyte development, mitophagy is also critical for the programmed elimination of sperm-contributed paternal mitochondria from fertilised oocytes during embryogenesis in *Caenorhabditis elegans* [9–11]. However, it is not clear as to whether this is also true in mammals [12–14]. Similarly, mitophagy is required for developmentally induced enterocyte cell death which occurs in the midgut of *Drosophila melanogaster* during metamorphosis [15].

Metabolic reprogramming is a characteristic of many differentiation pathways, and mitophagy has recently been described to influence this process. Programmed mitophagy has been shown to direct the

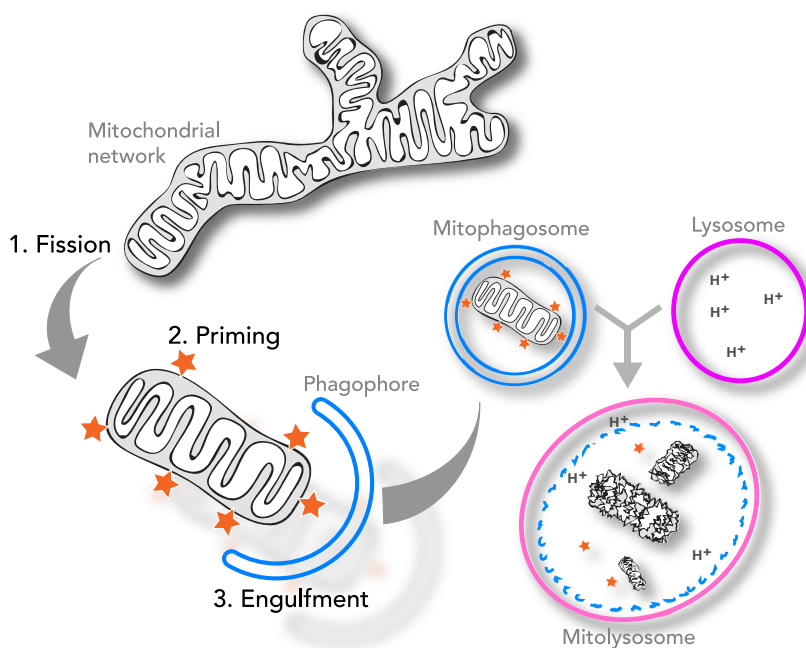


Fig. 1. The requirements for mitophagy. Cartoon depicting the molecular events occurring during mitophagy. (1) Fission of the mitochondrial network into individual mitochondrion/mitochondrial fragments; (2) Marking of these to prime them for recognition by the autophagy machinery; (3) Formation of the autophagic phagophore to engulf the primed mitochondrion. Once engulfed, the mitochondrion-containing autophagosome (mitophagosome) fuses with a lysosome to form the mitolysosomes where degradation and recycling occurs.

metabolic transition which occurs during cardiomyocyte maturation. During the neonatal period, cardiomyocyte substrate preference switches from carbohydrates to fatty acids, and this is concurrent with the replacement of small, fetal mitochondria with morphologically distinct mature, adult mitochondria better suited to support the heightened energetic demand of mature cardiac muscle [16]. Remarkably, mitophagy has been implicated in this maturation process: genetic disruption of mitophagy-related signalling in mice results in fetal-like mitochondria in adult animals, the development of progressive cardiomyopathy and death by 7–8 weeks of age [17].

Another example where developmentally induced mitochondrial changes occur in mitochondria is during the conversion of myoblasts into myotubes during skeletal muscle fibre development. Upon differentiation, when myoblasts ultimately fuse into multinucleated myofibres, their metabolic profile is reconfigured from glycolysis towards oxidative phosphorylation (OXPHOS) [18]. Studies using cultured murine C2C12 myoblasts have shown that at this time, the mitochondrial population is initially cleared and then replenished with a structurally distinct network [19]. Mitophagy is believed essential for this process and is, therefore, a potential key regulator of myogenesis [19].

Intriguingly, mitophagy itself can also promote glycolysis, a phenomenon first reported in cancer cells which can enhance mitophagy to upregulate glycolysis during prolonged mitotic arrest [20]. A mitophagy-triggered metabolic switch also occurs during the development of the embryonic mouse retina [21]. In retinal ganglion cells (RGCs), the first retinal cell type to arise, differentiation occurs alongside a switch from OXPHOS to a highly glycolytic state. A block in mitophagy abrogates RGC differentiation and the associated bioenergetic changes, highlighting the pivotal role of mitophagy in this process [21].

Further elaborating on the fundamental importance of mitophagy, it has emerged as a key determinant for the maintenance of stem cell identity in embryonic stem cells, hematopoietic stem cells, and satellite cells, a population of muscle stem cells required for regeneration of skeletal muscle [22–24]. Mitophagy is also an important feature of somatic cell reprogramming to induce pluripotent stem cells (iPSCs). Somatic cells and iPSCs have distinct metabolic identities, whereby somatic cells rely predominantly on OXPHOS and iPSCs are highly glycolytic. This metabolic shift is essential for reprogramming efficiency and is accompanied by a characteristic reduction in mitochondrial content [25]. Critically, mitophagic activity accounts for the decrease in mitochondrial

mass observed at the onset of somatic cell reprogramming and contributes to successful completion of iPSC generation [22,26,27].

Mechanistic basics for mitophagy initiation

Mitophagy is a mechanistically elaborate process and critically depends on the regulated convergence of multiple signalling events. The basic requirements for mitophagy are summarised in Fig. 1. These include (a) fission of the mitochondrial network to produce a suitably sized mitochondrion that can be readily engulfed by the forming autophagosome; (b) priming of the mitochondrion for recognition by the autophagy machinery; and (c) engulfment of the marked mitochondrion by the forming autophagosome. Whether these three steps occur independently and sequentially, or in combination, remains to be determined.

Fission

Mitochondrial dynamics are integral to mitochondrial function and the mechanics of mitophagy. Size constraints dictate that a typical mitochondrion cannot be accommodated within an autophagosome unless the network undergoes significant structural alteration. Mitochondrial fission refers to the severing of the mitochondrial network into smaller portions and is catalysed by the GTPase dynamin-related protein 1 (Drp1) [28]. At sites where endoplasmic reticulum (ER) tubules contact mitochondria, Drp1 is recruited from the cytosol by several adaptor proteins located at the surface of the organelle [29–31]. Subsequent Drp1 oligomerisation and GTP hydrolysis leads to complete membrane constriction at fission sites [32]. Importantly, fission commonly precedes and is necessary for mitophagy. Random fission events have been observed to generate two mitochondrial daughter units with uneven membrane potential [4]. The fragments with diminished, unrecoverable membrane potential were unlikely to rejoin the mitochondrial network and more prone to turnover, thus providing a means to ensure the overall quality of the mitochondrial network [4]. Conversely, it has also been shown that starvation-induced elongation of the mitochondrial network spares mitochondria from degradation, enhancing OXPHOS potential during energy deprivation [33,34]. Enhanced mitochondrial connectivity has also been observed when mitochondrial OXPHOS is upregulated [35]. Under these conditions, mitophagy is blocked as a direct consequence of defective mitochondrial fission [36].

It is essential to consider how the signals coordinating mitochondrial fission and mitophagy may be coupled. A direct role for the key mitophagy initiating kinase PTEN-induced putative kinase 1 (PINK1), which is discussed in more detail below, has been proposed to play a role in this process. Following pharmacological agent-induced depolarisation, PINK1 activity was shown to alleviate inhibitory protein kinase A (PKA)-dependent Drp1 phosphorylation to promote fission [37]. This is due to the displacement of PKA from AKAP1, a mitochondrial scaffolding protein that recruits PKA to the outer mitochondrial membrane (OMM), by an unknown mechanism. Another mitochondrial and mitophagy-related protein, FUN-14 domain containing protein 1 (FUNDCl), has been shown to instigate fission through direct physical association with Drp1. Wu *et al.* [38] demonstrated that a shift to hypoxic conditions induces the relocation of FUNDCl to mitochondria-associated ER membranes (MAMs), where it binds the MAM protein calnexin. Longer exposure to hypoxia reduces the interaction between FUNDCl and calnexin, with the latter eventually being replaced with Drp1. Research by Chen *et al.* [39] showed that while Drp1 normally associates with the mitochondrial fusion factor OPA1, stress inducers such as selenite or carbonyl cyanide *p*-(trifluoromethoxy)-phenylhydrazon promote the binding of FUNDCl to Drp1 instead.

The energy-sensing kinase AMP-activated protein kinase (AMPK) has strong links to mitochondrial function and has also been connected to mitochondrial dynamics [40]. Upon its activation, either directly or through inhibition of the mitochondrial electron transport chain, AMPK phosphorylates mitochondrial fission factor (MFF) which increases mitochondrial Drp1 recruitment and subsequent fission. This is speculated to assist the mitophagic process.

Despite this, the mitophagy-specific requirement of Drp1 has not been comprehensively established and alternate fission mechanisms may exist. Much work on the role of Drp1 in mitophagy has employed the use of mDivi1, a Drp1 small molecule inhibitor [41]. Recent work suggests mDivi1 does not target Drp1 and instead functions as a mitochondrial complex I inhibitor [42]. Additionally, there is mounting evidence for Drp1-independent mitophagy [43–45]. Remarkably, research from the Dorn laboratory revealed that mitophagy is actually promoted in the absence of Drp1 and inhibited in the absence of the fusion proteins Mfn1 and Mfn2 [44]. While adding to the complexity of the situation, this nonetheless attests to the importance of mitochondrial dynamics to mitophagy. Elegant work from Yamashita *et al.* [45] demonstrated an alternative scheme for

mitochondrial fission upon mitophagy involving the autophagic machinery itself. They found that Drp1 is clearly absent from sites of autophagosome biogenesis at mitochondria and is dispensable for mitochondrial fission induced by a range of mitophagy stimuli. The autophagy machinery was first demonstrated to play a role in mitochondrial fission by the Chu Lab, which showed that siRNA of ATG7 or LC3 prevented mitochondrial fission in cells lacking PINK1 [46]. Interestingly, Drp1 was also implicated in this fission, implying that Drp1-dependent and autophagy-dependent fission may operate independently and co-operatively. However, the contexts of these fission events are far from clear, and it remains to be determined if different mitophagy pathways have differing mitochondrial fission requirements.

Priming mitochondria for mitophagy

The marking, or priming, of mitochondria for clearance is an essential step in mitophagy. Multiple factors, or receptors, have been identified which can orchestrate this by distinct mechanisms. Mitophagy receptors are central to mitochondrial priming and can be classified into two distinct groups; those which exist as integral mitochondrial proteins and those which indirectly associate with mitochondria through binding ubiquitylated mitochondrial proteins [47]. All known protein mitophagy receptors possess a short, linear LC3-interacting region (LIR) motif, which is responsible for their interaction with LC3/GABARAP proteins on the autophagosomal membrane. It consists of a core consensus sequence, as indicated by the single letter amino acid code: (W/F/Y)XX(L/I/V), where X can be any amino acid, and is often flanked by acidic residues [47]. The LIR was first characterised in the ubiquitin-binding protein p62/SQSTM1 as a critical determinant for the incorporation of polyubiquitylated aggregates into forming autophagosomes [48]. Since this work, multiple LIR motif-containing proteins have been identified and mediate the autophagy of various cargoes, in addition to mitochondria [49]. Whether these receptors rely solely on their LIR motifs for cargo incorporation is not clear, but the receptors themselves are critical.

PINK1/Parkin-mediated mitophagy

The most studied mitophagy pathway to date is mediated by the products of two genes that are mutated in autosomal recessive forms of Parkinson's disease (PD) [50,51]. Specifically, the mitochondrial-associated protein kinase PINK1 and cytosolic RBR E3-ubiquitin ligase Parkin play critical roles in damage-induced

mitophagy (see [3,52,53] for the landmark papers and reviewed in [54]). Under basal or steady-state conditions, cellular PINK1 levels remain low due to constitutive processing at mitochondria prior to subsequent N-end rule degradation [55]. PINK1 acts as a molecular sensor of mitochondrial damage; mitochondrial membrane depolarisation inhibits PINK1 cleavage and degradation, leading to its accumulation on the OMM. PINK1 stabilisation is associated with enhanced kinase activity, dimerisation and autophosphorylation [56–59]. The subsequent recruitment of cytosolic Parkin to mitochondria is dependent on PINK1 activation [52,59–62]. Parkin exists in an auto-inhibited conformation mediated by intra-molecular interactions within its UBL domain [63,64]. PINK1-mediated phosphorylation of both Parkin [57,65] and ubiquitin at their respective serine 65 residues is necessary for full Parkin activation [66–68]. Parkin catalyses the *de novo* conjugation and elongation of pre-existing ubiquitin chains onto a host of OMM proteins, which are predicted to serve as PINK1 substrates [66–68]. This increase in phospho-ubiquitin chains on depolarised mitochondria constitutes a feed-forward amplification loop that results in the completion of mitophagy [69–71]. It is important to note that Parkin is not the only E3 ubiquitin ligase that has been implicated in mitophagy, with ligases such as MUL1 possibly performing a similar role [14]. Additionally, ubiquitylation itself is reversible and deubiquitylases have thus been shown to be negative regulators of mitophagy [72].

Ubiquitylation of OMM proteins enables the recruitment of mitophagy receptors which possess a ubiquitin-binding domain in addition to a LIR motif. These include p62/SQSTM1, OPTN, NDP52, TAX1BP1 and NBR1 [73]. Utilisation of genome-editing technology has revealed that NDP52 and OPTN are the essential receptors for mitophagy, with a lesser contribution from TAX1BP1, while NBR1 and p62 are dispensable [74,75]. Surprisingly, OPTN and NDP52 are recruited to mitochondria even in the absence of Parkin and mitochondrial damage, and this is sufficient to induce mitophagy, albeit at a very low level [74].

Recently, the Levine laboratory identified prohibitin 2 (PHB2) as the first inner mitochondrial membrane (IMM) mitophagy receptor, which they found to be required for Parkin-dependent mitophagy [76]. In Parkin-overexpressing cells subject to mitochondrial damaging agents, proteasomal degradation of the OMM makes IMM components accessible. Successful mitophagy relies on an interaction between PHB2 and LC3 via its LIR [76]. Taken together, this suggests that a combination of mitophagy receptors is required for efficient mitochondrial priming.

Although much of the experimental evidence obtained to date supports the notion that PINK1 and Parkin are involved in damage-induced mitophagy, this pathway has also been implicated in the regulation of programmed mitophagy. Indeed, the previously mentioned mitophagy that occurs during cardiomyocyte development is described to be PINK1/Parkin-dependent [17].

BNIP3 and NIX/BNIP3L

Collectively, the findings amassed to date undoubtedly substantiate the importance of PINK1 and Parkin as central regulators of mitophagy. However, multiple other mitophagy pathways exist in addition. Indeed, our overall understanding of mitophagy has been propelled by evidence demonstrating that mitophagy is also mediated by receptors constitutively localised to mitochondria in a PINK1/Parkin-independent manner. The hypoxia-inducible proteins BNIP3L/NIX and its closely related homologue BNIP3 were the first mammalian mitophagy receptors identified [7,8,77]. They were first characterised as pro-apoptotic BH3-only Bcl-2 family proteins, both localised to the OMM by way of a C-terminal transmembrane domain [78]. Both NIX and BNIP3 contain identical LIRs at their N termini which facilitate interaction with autophagosomal LC3/GABARAP proteins [79]. Both NIX and BNIP3 have been implicated in mitophagy; however, currently NIX is the most widely studied of the two proteins.

The mitophagic role of NIX was first illustrated in the context of erythropoiesis. NIX, which is transcriptionally upregulated during erythroid differentiation, is essential for mitochondrial clearance via mitophagy during the terminal stages of this process [7,8,80]. Strikingly, in erythrocytes from NIX-deficient mice, mitochondria fail to be incorporated into autophagosomes, implying that NIX is critical for recruitment [7,8]. Ablation of NIX interrupts erythroid maturation, resulting in anaemia, reticulocytosis and erythroid-myeloid hyperplasia.

Besides erythroid differentiation, NIX is also the receptor required for mitophagy during somatic cell reprogramming to iPSCs and for RGC differentiation [21,26]. During RGC differentiation, local hypoxia transcriptionally induces NIX, accumulation of which triggers both mitochondrial removal and activation of glycolysis. Retinas from mice lacking NIX retain their mitochondria and exhibit fewer RGCs [21].

NIX is also involved in mitophagy induced when the demand for mitochondrial OXPHOS is high [81]. In these circumstances, the small GTPase Rheb is recruited to the OMM where it is proposed to

stimulate mitophagy through binding NIX and recruiting LC3 [81]. Accelerated rejuvenation of the mitochondrial pool is believed to contribute to increased OXPHOS efficiency.

A role for BNIP3 and mitophagy has been implicated in cancer development. In a mouse model of mammary tumorigenesis, loss of BNIP3 resulted in reduced mitophagy, the accumulation of functionally impaired mitochondria, elevated ROS production and consequent stabilisation of HIF1 α [82]. Increased HIF1 α target gene expression favoured angiogenesis and bioenergetic changes associated with Warburg metabolism that ultimately promoted invasiveness and metastasis.

Although BNIP3 and NIX are primarily under transcriptional control, their mitophagic activity is also activated by post-translational modification. Phosphorylation by an unknown kinase adjacent to the LIR of NIX and BNIP3 strengthens the association with LC3 to enhance mitophagy [83,84].

FUNDC1

FUNDC1, an OMM-spanning protein with an N-terminal LIR motif, is another mitophagy receptor which functions independently of the PINK1/Parkin axis, and is responsible for hypoxia-induced mitophagy [85]. Short-term regulation of FUNDC1 is carefully controlled by its phosphorylation status. In unstressed conditions, FUNDC1 is phosphorylated by Src kinase at tyrosine 18 and CK2 at serine 13, blocking its interaction with LC3 [85,86]. PGAM5 is a mitochondrially localised phosphatase capable of dephosphorylating FUNDC1 at serine 13 [86]. Its activity is usually held in check through inhibitory interaction with Bcl-xL; however, Bcl-xL is degraded upon hypoxic stress, thereby enhancing PGAM5 phosphatase activity and promoting the targeting of mitochondria to LC3-decorated autophagosomes for engulfment [87]. The interaction between FUNDC1 and LC3 can be further strengthened following UNC-51-like kinase 1 (ULK1)-dependent phosphorylation of FUNDC1 at serine 17 [88]. Beyond phosphorylation, ubiquitylation has also been implicated as a FUNDC1 control mechanism [89]. The OMM-localised E3 ligase MARCH5 can ubiquitylate FUNDC1 at the early stages of hypoxia, before FUNDC1 dephosphorylation, to prevent excessive mitophagy [89].

Bcl2L13

Bcl2L13, first identified as a pro-apoptotic and atypical Bcl2 family member [90], is another

OMM-anchored mitophagy receptor which can recruit LC3/GABARAP proteins to mitochondria during mitophagy [43]. Interestingly, this study showed that Bcl2L13 is multifunctional with respect to mitophagy as it mediates both mitochondrial fission and mitophagy upon mitochondrial damage. While the BH2 domains of Bcl2L13 were required for mitochondrial fragmentation, mutation of its internal LIR motif precludes association with LC3. Thus, both BH2 and LIR domains appear vital for mitophagy.

FKBP8

FKBP8/FKBP38 is a unique member of the FK506 family of binding proteins, distinguished by the fact that it possesses Ca²⁺/calmodulin-activated peptidyl-prolyl cis-trans isomerase activity. It has been linked to multiple cellular processes including apoptosis and regulation of mTOR [91]. Among the known mitophagy receptors in the OMM, FKBP8 is unique in that it itself avoids mitophagic turnover by rerouting to the ER [92,93]. While solid evidence of endogenous FKBP8 participating in mitophagy is lacking, it has been demonstrated that Parkin-independent mitophagy ensues upon overexpression of both LC3 and FKBP8. The exact mechanism by which FKBP8 escapes degradation is unclear, as are the reasons for this intriguing behaviour, although it depends on a number of basic residues within its C-terminal sequence [92,93].

Cardiolipin

Protein receptors are not the only mode of mitophagy priming. The mitochondrially localised and unique anionic phospholipid cardiolipin (CL) can also act as a receptor. CL is unusual in that it is almost exclusively confined to the IMM. Here, it is an important regulator of mitochondrial bioenergetics, as association between CL and electron transport chain components is required for their optimal activity [94]. Exposure to a range of mitophagic stimuli has also been shown to cause translocation of CL to the OMM, where it is free to interact with the N-terminal of LC3 to facilitate capture of the mitochondrion into the forming autophagosome [95]. The translocation of CL to the OMM was recently shown to require NDPK-D, an intermembrane space-localised nucleoside diphosphate kinase, which binds CL following mitophagy induction [96]. CL has also been shown to regulate mitochondrial fission and fusion through recruitment and activation of Drp1 and OPA1; therefore, it may play multiple roles in mitophagy [97–99]. Additionally, other lipids may also play an important role in regulating mitophagy [100].

While we have discussed the various mitochondrial receptors individually, it is possible that they do not operate in isolation. For example, there is evidence to support the idea that crosstalk exists between PINK1/Parkin-mediated mitophagy and OMM receptor-mediated mitophagy pathways [101–103]. However, it has been clearly demonstrated that NIX-mediated mitophagy is active even in the absence of a functional PINK1/Parkin pathway, as overexpression of NIX in human fibroblasts, isolated from patients with Parkin or PINK1-related early-onset PD, was capable of restoring carbonyl cyanide *m*-chlorophenylhydrazone-induced mitophagy [104]. While potential crosstalk cannot be ruled out, its nature and regulation require further investigation.

Mitochondrial engulfment by the autophagy machinery

Coincident with marking mitochondria for degradation, the autophagy machinery must be activated to drive formation of the engulfing autophagosome. While the relevant upstream signalling events are well defined in the context of classical starvation-induced autophagy (reviewed in [105–107]), the sensing mechanisms and signals that set the stage for targeted mitophagy induction are less clear.

Autophagy activating signals

At the helm of the canonical starvation-induced autophagy pathway is the ULK1 protein kinase complex (comprising ULK1, ATG13, FIP200 and ATG101). ULK1 complex activation is essential for the early stages of autophagosome biogenesis [105] and as such has been implicated in both programmed and damage-induced mitophagy. With respect to the former, ULK1 knockout (KO) mice display impaired removal during reticulocyte development [108,109]. As for the latter, ULK1 has also been shown to be critical for mitochondrial clearance in the PINK1/Parkin-mediated pathway [74,110] as well as during hypoxia [88].

mTORC1-ULK1

The major negative regulator of canonical autophagy is the nutrient-sensing protein kinase complex mammalian target of rapamycin complex 1 (mTORC1). In nutrient replete conditions, active mTORC1 restrains autophagic induction through inhibitory phosphorylation of ULK1 [105]. Repression of mTORC1 and dephosphorylation of ULK1 is essential for the activation of autophagy. In terms of mitophagy, there has

been only limited analysis of the mTORC1 signalling pathway. In cybrid cells depleted of mtDNA, but not in wild-type counterparts, mitophagy is provoked using the mTORC1 inhibitor rapamycin in conjunction with mitochondrial depolarisation [111]. In line with this, during neurogenesis, rapamycin treatment has also been observed to induce mitophagy and promote RGC differentiation [21]. Conflictingly, mitophagy mediated by Rheb under OXPHOS-dependent conditions occurs in the absence of mTORC1 deactivation [81]. Further work is obviously needed to clarify the role of the ULK1-mTORC1 axis during mitophagy stimulation.

AMPK-ULK1

Along with mTORC1, AMPK plays a significant role in regulating ULK1 activity in general autophagy. Multiple studies have identified AMPK-dependent phosphorylation sites within ULK1 [112–116]. However, with varying overlap in sites identified between studies, and both activating and inhibitory effects reported, the relationship between AMPK and ULK1 remains obscure.

Consistent with a functional connection between AMPK and ULK1, mitochondrial removal is suppressed during the terminal differentiation of erythrocytes from AMPK α 1 KO mice [117]. It has also been reported that upon hypoxia, AMPK directly phosphorylates ULK1 at S555 [118]. This triggers ULK1 translocation to mitochondria and is critical for mitophagic induction. As mentioned previously, AMPK has been shown to regulate mitochondrial fission [40] and this opens up the possibility that AMPK coordinates multiple steps of mitophagy induction.

Recruitment of the autophagy machinery by mitophagy receptors

The long-held notion is that a key function of the mitophagy receptors is to recruit a forming autophagosome to the mitochondrion via their ability to bind the autophagosomal LC3/GABARAP proteins. However, this may not be the only role the receptors play, with evidence suggesting that they may function earlier at the autophagosome initiation stage. Autophagosome generation is believed to be initiated at the mitochondrion itself and mitochondrial recruitment of upstream autophagy factors, including ULK1, DFCP1 and ATG14, has been shown to occur in the absence of LC3 [45,110]. Indeed, during mitophagy, there does not seem to be an absolute requirement for LC3/GABARAP proteins in autophagosome formation

[119,120]. Instead, recent observations suggest that LC3 functions at the autophagosome-lysosome fusion step and can mediate the subsequent degradation of the inner autophagosomal membrane [119,120]. Therefore, receptor interaction with the autophagy signalling machinery may be key.

In the case of PINK1/Parkin-dependent mitophagy, recruitment of the autophagy machinery to mitochondria has been reported to occur downstream of mitochondrial priming. Although ULK1 regulation by AMPK and mTORC1 occurs independently to the core mitophagy receptors OPTN and NDP52, they are

thought to be indispensable for ULK1 recruitment to mitochondria [74]. The subsequent recruitment of LC3, WIPI1 and DFCEP1 is thus also reliant on both OPTN and NDP52.

Notably, FUNDC1 was also proposed to be a mitochondrial receptor for ULK1 [88]. In this hypoxia-induced mitophagy pathway, ULK1 translocation to mitochondria is dependent on FUNDC1, and their association is necessary to accomplish mitochondrial degradation [88].

As is summarised in Table 1, a complex molecular picture of mitophagy is emerging with multiple

Table 1. Proteins involved in the different stages of mitophagy induction.

Protein	Function	References
Stage of mitophagy		
Fission		
Drp1	Dynamin-related GTPase	[37–40,46]
PINK1	Mitochondrial protein kinase	[37]
PKA	Multifunctional protein kinase	[37]
AKAP	Mitochondrial PKA scaffolding protein	[37]
AMPK	Energy-sensing protein kinase	[40]
MFF	DRP1-interacting mitochondrial protein	[40]
FUNDC1	Mitophagy receptor	[38,39]
BCL2L13	Mitophagy receptor	[43]
FIP200/ATG14/WIPI2/ATG7	Autophagy initiation machinery	[45,46]
CL	Mitochondrial phospholipid	[96–98]
Priming		
PINK1	Mitochondrial protein kinase	[52–54]
Parkin	E3 ubiquitin ligase	[3,54]
Phospho-ubiquitin	PINK1 substrate	[66–68]
p62	Ubiquitin-binding autophagy/mitophagy receptor	[73–75]
OPTN	Ubiquitin-binding autophagy/mitophagy receptor	[73–75]
NDP52	Ubiquitin-binding autophagy/mitophagy receptor	[73–75]
NBR1	Ubiquitin-binding autophagy/mitophagy receptor	[73,74]
TAX1BP1	Ubiquitin-binding autophagy/mitophagy receptor	[73,74]
PHB2	Mitophagy receptor	[76]
NIX	Mitophagy receptor	[7,8,21,26,81]
Rheb	mTOR-activating GTPase	[81]
BNIP3	Mitophagy receptor	[82,83]
FUNDC1	Mitophagy receptor	[85–89]
PGAM5	Mitochondrial protein phosphatase	[86,87]
CK2	Protein kinase	[86]
Src	Tyrosine protein kinase	[85,86]
MARCH5	Mitochondrial E3 ubiquitin ligase	[89]
ULK1	Autophagy-initiating protein kinase	[88]
BCL2L13	Mitophagy receptor	[43]
FKBP8	Mitophagy receptor	[92,93]
CL	Mitochondrial phospholipid	[95,96]
Engulfment		
ULK1	Autophagy-initiating protein kinase	[73,74,88,108–110]
mTORC1	Nutrient-sensing protein kinase	[21,81,111]
AMPK	Energy-sensing protein kinase	[112–116]
OPTN	Mitophagy receptor	[73]
NDP52	Mitophagy receptor	[73]
FUNDC1	Mitophagy receptor	[88]

mechanisms proposed to regulate both damage-induced and programmed mitophagy. However, most of these mechanistic insights have been derived from cultured cell lines treated under nonphysiological conditions. Therefore, the major future challenge is to determine when and where these pathways operate *in vivo*.

Investigating mammalian mitophagy *in vivo*

It has proved challenging to evaluate the physiological significance of numerous *in vitro* observations pertaining to mitophagy. This was due to a distinct lack of tools that facilitate the reliable detection of mammalian mitophagy *in vivo*. Recently, two mouse models have been described that utilise fluorescent reporter systems to enable the detection of mitophagy *in vivo* (*mito-QC* reported by [121] and *mt-Keima* reported by [122]). These models exploit the acid labile properties of fluorescent reporter proteins to provide end-point readouts of mitophagy *in vivo*. As such, the readout of mitophagy in the context of the current *in vivo* reporters is defined in terms of mitochondrial delivery to the acidic microenvironment of the lysosome. Under these conditions, the acidity either quenches the GFP signal (in the case of *mito-QC*) or shifts the excitation wavelength (in the case of *mt-Keima*, see Fig. 2). Both reporters faithfully monitor mitophagy and are ubiquitously expressed, though from different loci (*Rosa26* for *mito-QC* and *Hip11* for *mt-Keima*). As the reporters monitor the end-point of mitophagy, they do not distinguish between distinct mitophagic pathways. However, the reporters localise to different mitochondrial compartments: mitochondrial outer membrane localisation for *mito-QC* versus matrix localisation for *mt-Keima*. This localisation is important as it may help distinguish between the previously mentioned MDV pathway, which represents an alternate mode of mitochondrial protein delivery to lysosomes [1]. In contrast to *mt-Keima*, the MOM topology of *mito-QC* means that it will never be delivered to the lysosomal lumen through the more classical transport mechanism of the MDV pathway – hence, the mCherry-only signal cannot be a consequence of this. Another model termed MitoTimer has also been described [123]; however, this is an inducible model designed to monitor mitochondrial biogenesis over time, as well as mitophagy. Differences between these models are illustrated in Fig. 2, and these have also been recently reviewed [124]. Regardless, these collective advances have transformed the landscape of mammalian mitophagy, enabling the interrogation of mitophagy and

mitochondrial homeostasis in a variety of physiological contexts [54,125].

Basal mitophagy as a pervasive yet heterogeneous process in tissues

Perhaps, the most surprising conceptual discovery arising from these models is converging data that demonstrate the basal nature of mitophagy *in vivo*. Although differences in these models are well described [121], both studies report highly pervasive instances of mitophagy under normal everyday conditions. This is in stark contrast to the still widely held perception of mitophagy as merely an ‘activated autophagy response’ triggered to mitigate mitochondrial meltdown. This raises several important and unanswered questions: what are the tissue-specific regulators of basal mitophagy *in vivo*? Do any of the aforementioned effectors of stimulus-induced mitophagy modulate basal homeostatic mitophagy *in vivo*? Does disruption of these basal pathways have any pathological consequences?

Basal mitophagy and metabolic context

Although both mitophagy models revealed an extensive amount of basal mitophagy, studies using *mito-QC* have highlighted the striking heterogeneity in mitophagy levels between defined cellular subsets *in vivo*. Thus, while all organ systems exhibit some degree of mitochondrial turnover, not all cells exhibit comparable levels of mitophagy *in vivo*. This is particularly evident in organ systems of high metabolic demand such as the nervous system, the renal system and cardiovascular system. In the brain, Sun *et al.* [122] were able to demonstrate regionalised instances of mitophagy in unlabelled tissue. However, given the breathtaking morphological complexity and diversity of neural subtypes, a more precise analysis is required to accurately interpret such observations in anisotropic tissue. Using *mito-QC* combined with immunohistochemistry and high-resolution microscopy, McWilliams *et al.* [121] showed specifically that labelled cerebellar Purkinje neurons exhibit robust levels of mitophagy, which was largely enriched within the somata. Future work will be essential to determine the spatial profile and regulation of mitophagy in different neuronal and nonneuronal subtypes. Furthermore, could basal mitophagy be elevated or diminished in genetic models of neurodegenerative disease? Interestingly, Sun *et al.* demonstrated diminished mitophagy in the context of Huntingtin overexpression [122]. PINK1/Parkin signalling has gained much attention in stimulated

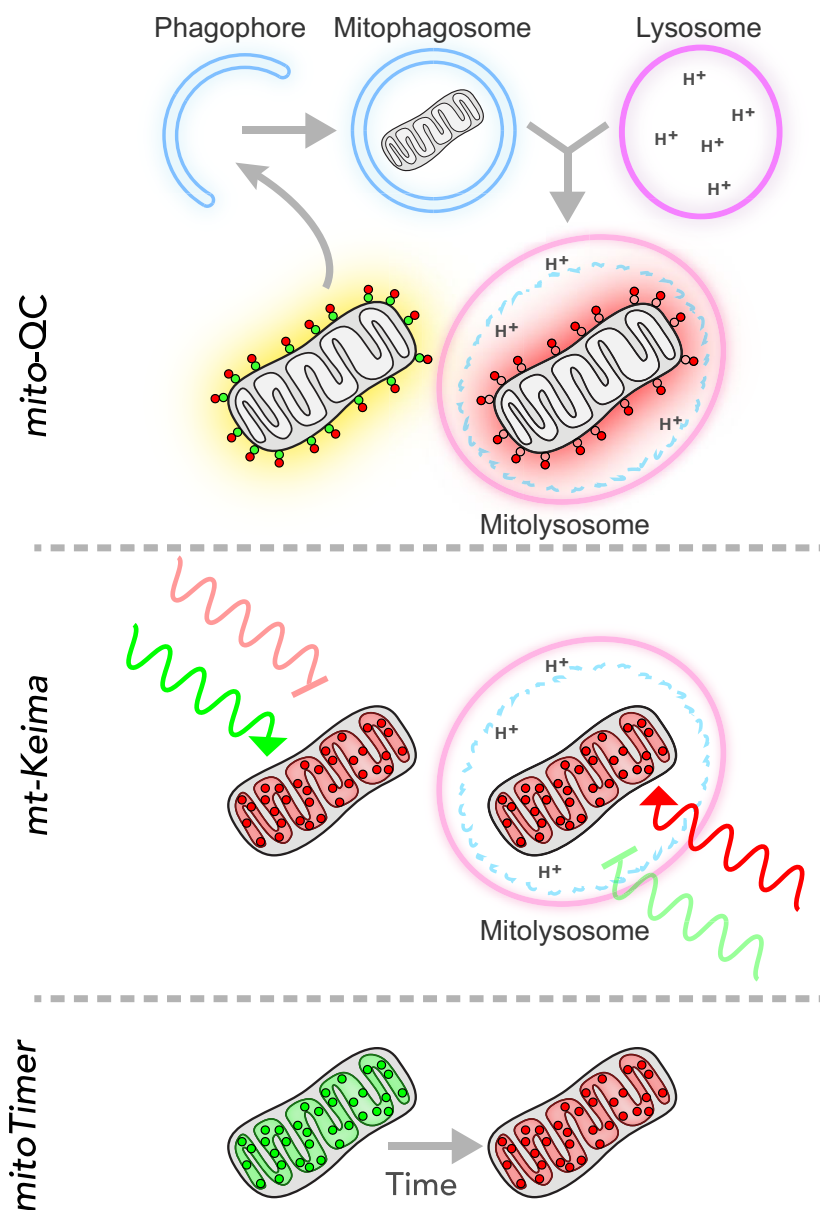


Fig. 2. Mitophagy reporters. Schematic detailing the different reporters used to monitor mitophagy. In *mito-QC*, a tandem mCherry-GFP tag is targeted to the OMM. Under normal cytosolic conditions the mitochondria display both mCherry and GFP fluorescence. However, upon mitophagy, the low pH of the mitochondrion-containing lysosome (mitolysosome) is sufficient to quench the GFP, but not mCherry, signal. In *mt-Keima*, monomeric Keima protein is targeted to the mitochondrial matrix. Under normal cytosolic conditions, *mt-Keima* is excited by light peaking at 400 nm, whereas this increases to 568 nm under lysosomal conditions, i.e. upon mitophagy. *mt-Keima*'s emission spectra remains the same, at 620 nm, regardless of this pH shift. For *MitoTimer*, the fluorophore DsRed1-E5 was targeted to the mitochondrial matrix. Though strictly not designed to monitor mitophagy, it provides a powerful way in which to monitor the age of mitochondria as fluorescence shifts from green to red over a period of 48 h.

mitophagy pathways; however, we have no idea if this signalling influences basal mitophagy *in vivo*. Interrogating this signalling is also troublesome, due to lack of reliable tools available to robustly detect endogenous PINK1 protein at basal levels *in vivo*. The requirement of endogenous Parkin under conditions of chronic mitochondrial stress was elegantly demonstrated by the Youle laboratory, who observed progressive age-dependent DA neurodegeneration in Parkin KO mice crossed with the mutator (*Polg^{D257A}*) mouse model [126]. However, it remains unclear if dysregulated mitophagy lies at the root of pathology in the double-mutant mice. Understanding mitophagy in

a mouse model exhibiting symptomatic neurodegeneration will prove vital to the interpretation of dysregulated mitophagy and its contribution to the progression of diseases such as PD.

Outside of the nervous system, renal tubules exhibit a high metabolic demand. This requirement is in large part to drive the massive salt and protein reabsorption from the filtrate. As a consequence of this, these post-mitotic tubular cells are second only to heart cardiomyocytes in terms of their mitochondrial content [127]. It is, therefore, not surprising that impaired mitophagy has been postulated to contribute to a range of renal pathologies, including acute kidney

injury [128,129]. It should not be overlooked that exactly 60 years ago, scientists described perhaps the first instances of mitophagy by observing degraded mitochondria within lysosome-like structures in rabbit kidney tubules [130]. In wild-type *mito-QC* animals, a striking level of mitophagy was observed throughout the cortex of the adult kidney (see Fig. 3 and [121]). However, a clear difference in the degree of mitophagy was observed between distinct tubules of the nephron. Proximal convoluted tubules (PCTs) exhibited a major enrichment in mitophagy compared with distal convoluted tubules (DCTs), even though both types of tubule are loaded with mitochondria. This discovery highlighted another important conceptual advance: that the degree of mitophagy within a cell is independent from its mitochondrial mass. What then might account for the heterogeneity of this basal mitophagy

observed *in vivo*? Metabolic context is likely to play important role. For example, differences in adult renal mitophagy could be a function of altered metabolic states between tubules and/or functional demand that requires a constant turnover of mitochondria. Indeed, oxygen consumption is believed to be greater in PCTs versus DCTs [131]. It could also be that PCT mitochondria are more vulnerable to mitochondrial stress than their DCT counterparts, although this is highly speculative and the actual comparative mitochondrial biology between these structures remains to be elaborated in greater mechanistic detail. Nonetheless, comparative mitochondrial differences between tubules have been described. For example, mitochondrial membrane potential is reported to be different between DCTs and PCTs [132]. The same study reports higher ROS production in PCTs than DCTs. Taken together,

Adult kidney cortex

Nuclei Mitochondria Mitolysosomes

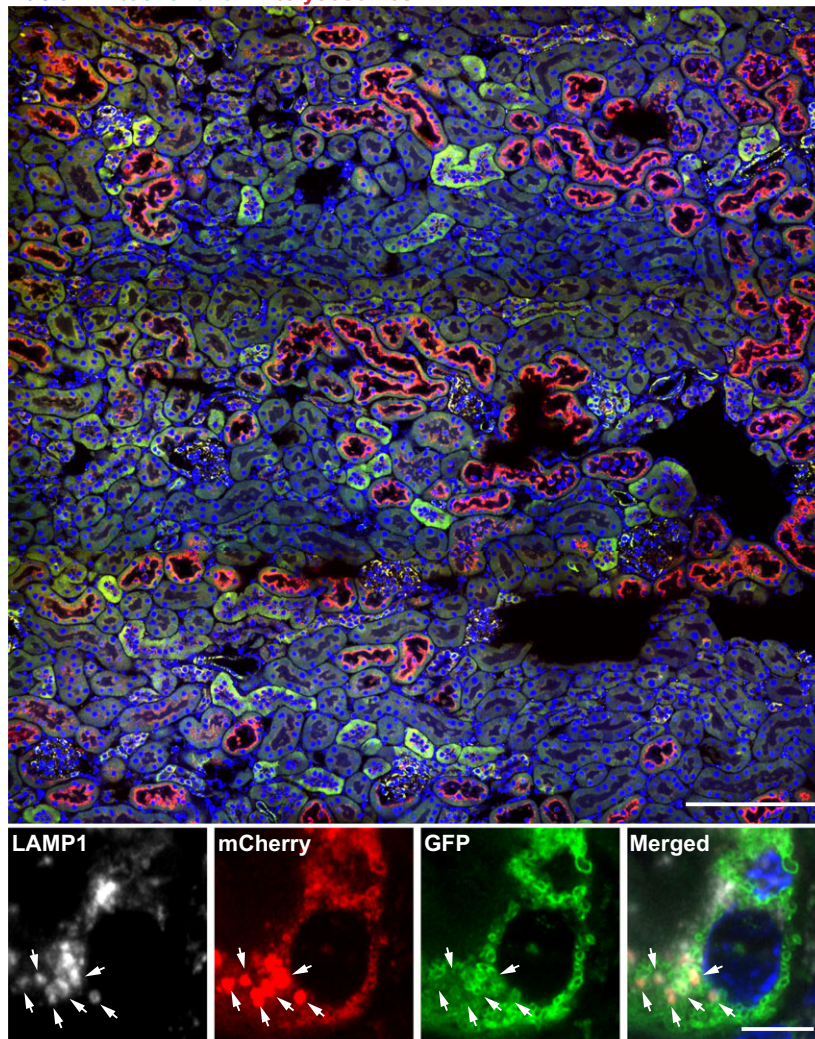


Fig. 3. Mitophagy in the cortex of the adult kidney. Upper panels show a confocal micrograph (tile scan) of a section of adult kidney cortex from the *mito-QC* reporter mouse under basal conditions. Note the distinct and polarised degree of mitophagy, as visualised by the ring-like mCherry (red) signals, in a subset of kidney tubules (PCTs). Scale bar, 200 μ m. Lower panels show a magnified *mito-QC* PCT cell detailing the GFP and mCherry mitochondrial signals. Arrows highlight mCherry-only signal co-localising with LAMP1, a marker for lysosomes, confirming mitophagy. Scale bar, 5 μ m. For details see McWilliams *et al.* [121].

these collective observations highlight the need for a deeper understanding of the interplay between mitochondrial metabolism and mitophagy within different cell subtypes *in vivo*.

The selective nature of autophagy *in vivo* is also highlighted when evaluating *mito*-QC data in the context of pioneering studies by Mizushima on *in vivo* autophagy [133]. Elegant work using a general autophagy reporter (GFP-LC3) mouse model showed elevated LC3-positive autophagosomes in adult kidney glomeruli, indicating elevated general autophagy in this renal structure. However, in *mito*-QC animals, low levels of mitophagy were observed in adult glomeruli compared with tubules. Unexpectedly, during embryonic development, this distinct renal mitophagy is reversed, with elevated mitochondrial turnover occurring in glomeruli but not tubules [121]. Therefore, not only are the physiological instances of autophagy highly context dependent but also demonstrate remarkable specificity, in this case for mitophagy.

Mitophagy modalities: intracellular and extracellular destruction

Autophagy has been classically studied as an intracellular, self-catabolic quality control pathway. However, it is emerging that this assumption may not be entirely accurate. Davis *et al.* [134] reported the trans-cellular elimination of mitochondria *in vivo*. They observed that neuronal mitochondria targeted for elimination were expelled from axonal evulsions within the optic nerve head and degraded by neighbouring glial cells. It will be important to understand and further elaborate how 'axonal-trans mitophagy' and other extracellular mitochondrial trafficking mechanisms contribute to mitochondrial homeostasis *in vivo*. These data challenge our assumptions of mitochondrial quality control pathways as being intrinsic to a particular cell and highlight the importance of studying mitochondrial homeostasis in the context of the neuroglial unit – a fundamental entity of the nervous system *in vivo* [54].

The future

Taken together, these discoveries demonstrate that further research is required to understand the orchestration of metabolism and mitophagy and their contribution to tissue development and long-term function. The molecular regulation of these processes remains enigmatic. It is, thus, accurate and appropriate to conclude that *in vivo* mitophagy remains a complex, context-dependent process: the next frontier of mitophagy research.

Acknowledgement

This work was supported by the Medical Research Council, UK (MC_UU_12016/4).

Author contributions

CER, TGM and IGG wrote the review.

References

- 1 Sugiura A, McLelland GL, Fon EA & McBride HM (2014) A new pathway for mitochondrial quality control: mitochondrial-derived vesicles. *EMBO J* **33**, 2142–2156.
- 2 Ashrafi G & Schwarz TL (2013) The pathways of mitophagy for quality control and clearance of mitochondria. *Cell Death Differ* **20**, 31–42.
- 3 Narendra D, Tanaka A, Suen DF & Youle RJ (2008) Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *J Cell Biol* **183**, 795–803.
- 4 Twig G, Elorza A, Molina AJ, Mohamed H, Wikstrom JD, Walzer G, Stiles L, Haigh SE, Katz S, Las G *et al.* (2008) Fission and selective fusion govern mitochondrial segregation and elimination by autophagy. *EMBO J* **27**, 433–446.
- 5 Ney PA (2015) Mitochondrial autophagy: origins, significance, and role of BNIP3 and NIX. *Biochim Biophys Acta* **1853**, 2775–2783.
- 6 Ney PA (2011) Normal and disordered reticulocyte maturation. *Curr Opin Hematol* **18**, 152–157.
- 7 Schweers RL, Zhang J, Randall MS, Loyd MR, Li W, Dorsey FC, Kundu M, Opferman JT, Cleveland JL, Miller JL *et al.* (2007) NIX is required for programmed mitochondrial clearance during reticulocyte maturation. *Proc Natl Acad Sci USA* **104**, 19500–19505.
- 8 Sandoval H, Thiagarajan P, Dasgupta SK, Schumacher A, Prchal JT, Chen M & Wang J (2008) Essential role for Nix in autophagic maturation of erythroid cells. *Nature* **454**, 232–235.
- 9 Al Rawi S, Louvet-Vallee S, Djeddi A, Sachse M, Culetto E, Hajjar C, Boyd L, Legouis R & Galy V (2011) Postfertilization autophagy of sperm organelles prevents paternal mitochondrial DNA transmission. *Science* **334**, 1144–1147.
- 10 Sato M & Sato K (2011) Degradation of paternal mitochondria by fertilization-triggered autophagy in *C. elegans* embryos. *Science* **334**, 1141–1144.
- 11 Zhou Q, Li H & Xue D (2011) Elimination of paternal mitochondria through the lysosomal degradation pathway in *C. elegans*. *Cell Res* **21**, 1662–1669.
- 12 Luo SM, Ge ZJ, Wang ZW, Jiang ZZ, Wang ZB, Ouyang YC, Hou Y, Schatten H & Sun QY (2013)

- Unique insights into maternal mitochondrial inheritance in mice. *Proc Natl Acad Sci USA* **110**, 13038–13043.
- 13 Song WH, Yi YJ, Sutovsky M, Meyers S & Sutovsky P (2016) Autophagy and ubiquitin-proteasome system contribute to sperm mitophagy after mammalian fertilization. *Proc Natl Acad Sci USA* **113**, E5261–E5270.
 - 14 Rojansky R, Cha MY & Chan DC (2016) Elimination of paternal mitochondria in mouse embryos occurs through autophagic degradation dependent on PARKIN and MUL1. *Life* **5**, e17896.
 - 15 Chang TK, Shrivage BV, Hayes SD, Powers CM, Simin RT, Wade Harper J & Baehrecke EH (2013) Uba1 functions in Atg7- and Atg3-independent autophagy. *Nat Cell Biol* **15**, 1067–1078.
 - 16 Porter GA Jr, Hom J, Hoffman D, Quintanilla R, de Mesy Bentley K & Sheu SS (2011) Bioenergetics, mitochondria, and cardiac myocyte differentiation. *Prog Pediatr Cardiol* **31**, 75–81.
 - 17 Gong G, Song M, Csordas G, Kelly DP, Matkovich SJ & Dorn GW 2nd (2015) Parkin-mediated mitophagy directs perinatal cardiac metabolic maturation in mice. *Science* **350**, aad2459.
 - 18 Wagatsuma A & Sakuma K (2013) Mitochondria as a potential regulator of myogenesis. *Sci World J* **2013**, 593267.
 - 19 Sin J, Andres AM, Taylor DJ, Weston T, Hiraumi Y, Stotland A, Kim BJ, Huang C, Doran KS & Gottlieb RA (2016) Mitophagy is required for mitochondrial biogenesis and myogenic differentiation of C2C12 myoblasts. *Autophagy* **12**, 369–380.
 - 20 Domenech E, Maestre C, Esteban-Martinez L, Partida D, Pascual R, Fernandez-Miranda G, Seco E, Campos-Olivas R, Perez M, Megias D *et al.* (2015) AMPK and PFKFB3 mediate glycolysis and survival in response to mitophagy during mitotic arrest. *Nat Cell Biol* **17**, 1304–1316.
 - 21 Esteban-Martinez L, Sierra-Filardi E, McGreal RS, Salazar-Roa M, Marino G, Seco E, Durand S, Enot D, Grana O, Malumbres M *et al.* (2017) Programmed mitophagy is essential for the glycolytic switch during cell differentiation. *EMBO J* **36**, 1688–1706.
 - 22 Liu K, Zhao Q, Liu P, Cao J, Gong J, Wang C, Wang W, Li X, Sun H, Zhang C *et al.* (2016) ATG3-dependent autophagy mediates mitochondrial homeostasis in pluripotency acquirement and maintenance. *Autophagy* **12**, 2000–2008.
 - 23 Ito K, Turcotte R, Cui J, Zimmerman SE, Pinho S, Mizoguchi T, Arai F, Runnels JM, Alt C, Teruya-Feldstein J *et al.* (2016) Self-renewal of a purified Tie2+ hematopoietic stem cell population relies on mitochondrial clearance. *Science* **354**, 1156–1160.
 - 24 Garcia-Prat L, Martinez-Vicente M, Perdiguero E, Ortet L, Rodriguez-Ubreva J, Rebollo E, Ruiz-Bonilla V, Gutarra S, Ballestar E, Serrano AL *et al.* (2016) Autophagy maintains stemness by preventing senescence. *Nature* **529**, 37–42.
 - 25 Folmes CD, Nelson TJ, Martinez-Fernandez A, Arrell DK, Lindor JZ, Dzeja PP, Ikeda Y, Perez-Terzic C & Terzic A (2011) Somatic oxidative bioenergetics transitions into pluripotency-dependent glycolysis to facilitate nuclear reprogramming. *Cell Metab* **14**, 264–271.
 - 26 Xiang G, Yang L, Long Q, Chen K, Tang H, Wu Y, Liu Z, Zhou Y, Qi J, Zheng L *et al.* (2017) BNIP3L-dependent mitophagy accounts for mitochondrial clearance during three factors induced somatic cell reprogramming. *Autophagy* **13**, 1543–1555.
 - 27 Ma T, Li J, Xu Y, Yu C, Xu T, Wang H, Liu K, Cao N, Nie BM, Zhu SY *et al.* (2015) Atg5-independent autophagy regulates mitochondrial clearance and is essential for iPSC reprogramming. *Nat Cell Biol* **17**, 1379–1387.
 - 28 Smirnova E, Griparic L, Shurland DL & van der Bliek AM (2001) Dynamin-related protein Drp1 is required for mitochondrial division in mammalian cells. *Mol Biol Cell* **12**, 2245–2256.
 - 29 Friedman JR, Lackner LL, West M, DiBenedetto JR, Nunnari J & Voeltz GK (2011) ER tubules mark sites of mitochondrial division. *Science* **334**, 358–362.
 - 30 Elgass KD, Smith EA, LeGros MA, Larabell CA & Ryan MT (2015) Analysis of ER-mitochondria contacts using correlative fluorescence microscopy and soft X-ray tomography of mammalian cells. *J Cell Sci* **128**, 2795–2804.
 - 31 Osellame LD, Singh AP, Stroud DA, Palmer CS, Stojanovski D, Ramachandran R & Ryan MT (2016) Cooperative and independent roles of the Drp1 adaptors Mff, MiD49 and MiD51 in mitochondrial fission. *J Cell Sci* **129**, 2170–2181.
 - 32 Ingerman E, Perkins EM, Marino M, Mears JA, McCaffery JM, Hinshaw JE & Nunnari J (2005) Dnm1 forms spirals that are structurally tailored to fit mitochondria. *J Cell Biol* **170**, 1021–1027.
 - 33 Gomes LC, Di Benedetto G & Scorrano L (2011) During autophagy mitochondria elongate, are spared from degradation and sustain cell viability. *Nat Cell Biol* **13**, 589–598.
 - 34 Rambold AS, Kostecky B, Elia N & Lippincott-Schwartz J (2011) Tubular network formation protects mitochondria from autophagosomal degradation during nutrient starvation. *Proc Natl Acad Sci USA* **108**, 10190–10195.
 - 35 Rossignol R, Gilkerson R, Aggeler R, Yamagata K, Remington SJ & Capaldi RA (2004) Energy substrate modulates mitochondrial structure and oxidative capacity in cancer cells. *Cancer Res* **64**, 985–993.
 - 36 MacVicar TD & Lane JD (2014) Impaired OPA1-dependent cleavage of OPA1 and reduced DRP1

- fission activity combine to prevent mitophagy in cells that are dependent on oxidative phosphorylation. *J Cell Sci* **127**, 2313–2325.
- 37 Pryde KR, Smith HL, Chau KY & Schapira AH (2016) PINK1 disables the anti-fission machinery to segregate damaged mitochondria for mitophagy. *J Cell Biol* **213**, 163–171.
- 38 Wu W, Lin C, Wu K, Jiang L, Wang X, Li W, Zhuang H, Zhang X, Chen H, Li S *et al.* (2016) FUNDC1 regulates mitochondrial dynamics at the ER-mitochondrial contact site under hypoxic conditions. *EMBO J* **35**, 1368–1384.
- 39 Chen M, Chen Z, Wang Y, Tan Z, Zhu C, Li Y, Han Z, Chen L, Gao R, Liu L *et al.* (2016) Mitophagy receptor FUNDC1 regulates mitochondrial dynamics and mitophagy. *Autophagy* **12**, 689–702.
- 40 Toyama EQ, Herzig S, Curchet J, Lewis TL Jr, Loson OC, Hellberg K, Young NP, Chen H, Polleux F, Chan DC *et al.* (2016) Metabolism. AMP-activated protein kinase mediates mitochondrial fission in response to energy stress. *Science* **351**, 275–281.
- 41 Cassidy-Stone A, Chipuk JE, Ingberman E, Song C, Yoo C, Kuwana T, Kurth MJ, Shaw JT, Hinshaw JE, Green DR *et al.* (2008) Chemical inhibition of the mitochondrial division dynamin reveals its role in Bax/Bak-dependent mitochondrial outer membrane permeabilization. *Dev Cell* **14**, 193–204.
- 42 Bordt EA, Clerc P, Roelofs BA, Saladino AJ, Tretter L, Adam-Vizi V, Cherok E, Khalil A, Yadava N, Ge SX *et al.* (2017) The putative Drp1 inhibitor mdivi-1 is a reversible mitochondrial complex I inhibitor that modulates reactive oxygen species. *Dev Cell* **40**, 583–594.e6.
- 43 Murakawa T, Yamaguchi O, Hashimoto A, Hikoso S, Takeda T, Oka T, Yasui H, Ueda H, Akazawa Y, Nakayama H *et al.* (2015) Bcl-2-like protein 13 is a mammalian Atg32 homologue that mediates mitophagy and mitochondrial fragmentation. *Nat Commun* **6**, 7527.
- 44 Song M, Mihara K, Chen Y, Scorrano L & Dorn GW 2nd (2015) Mitochondrial fission and fusion factors reciprocally orchestrate mitophagic culling in mouse hearts and cultured fibroblasts. *Cell Metab* **21**, 273–285.
- 45 Yamashita SI, Jin X, Furukawa K, Hamasaki M, Nezu A, Otera H, Saigusa T, Yoshimori T, Sakai Y, Mihara K *et al.* (2016) Mitochondrial division occurs concurrently with autophagosome formation but independently of Drp1 during mitophagy. *J Cell Biol* **215**, 649–665.
- 46 Dagda RK, Cherra SJ 3rd, Kulich SM, Tandon A, Park D & Chu CT (2009) Loss of PINK1 function promotes mitophagy through effects on oxidative stress and mitochondrial fission. *J Biol Chem* **284**, 13843–13855.
- 47 Birgisdottir AB, Lamark T & Johansen T (2013) The LIR motif - crucial for selective autophagy. *J Cell Sci* **126**, 3237–3247.
- 48 Pankiv S, Clausen TH, Lamark T, Brech A, Bruun JA, Outzen H, Overvatn A, Bjorkoy G & Johansen T (2007) p62/SQSTM1 binds directly to Atg8/LC3 to facilitate degradation of ubiquitinated protein aggregates by autophagy. *J Biol Chem* **282**, 24131–24145.
- 49 Khaminets A, Behl C & Dikic I (2016) Ubiquitin-dependent and independent signals in selective autophagy. *Trends Cell Biol* **26**, 6–16.
- 50 Kitada T, Asakawa S, Hattori N, Matsumine H, Yamamura Y, Minoshima S, Yokochi M, Mizuno Y & Shimizu N (1998) Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature* **392**, 605–608.
- 51 Valente EM, Abou-Sleiman PM, Caputo V, Muqit MM, Harvey K, Gispert S, Ali Z, Del Turco D, Bentivoglio AR, Healy DG *et al.* (2004) Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science* **304**, 1158–1160.
- 52 Matsuda N, Sato S, Shiba K, Okatsu K, Saisho K, Gautier CA, Sou YS, Saiki S, Kawajiri S, Sato F *et al.* (2010) PINK1 stabilized by mitochondrial depolarization recruits Parkin to damaged mitochondria and activates latent Parkin for mitophagy. *J Cell Biol* **189**, 211–221.
- 53 Narendra DP, Jin SM, Tanaka A, Suen DF, Gautier CA, Shen J, Cookson MR & Youle RJ (2010) PINK1 is selectively stabilized on impaired mitochondria to activate Parkin. *PLoS Biol* **8**, e1000298.
- 54 McWilliams TG & Muqit MM (2017) PINK1 and Parkin: emerging themes in mitochondrial homeostasis. *Curr Opin Cell Biol* **45**, 83–91.
- 55 Yamano K & Youle RJ (2013) PINK1 is degraded through the N-end rule pathway. *Autophagy* **9**, 1758–1769.
- 56 Aerts L, Craessaerts K, De Strooper B & Morais VA (2015) PINK1 kinase catalytic activity is regulated by phosphorylation on serines 228 and 402. *J Biol Chem* **290**, 2798–2811.
- 57 Kondapalli C, Kazlauskaitė A, Zhang N, Woodroof HI, Campbell DG, Gourlay R, Burchell L, Walden H, Macartney TJ, Deak M *et al.* (2012) PINK1 is activated by mitochondrial membrane potential depolarization and stimulates Parkin E3 ligase activity by phosphorylating Serine 65. *Open Biol* **2**, 120080.
- 58 Okatsu K, Oka T, Iguchi M, Imamura K, Kosako H, Tani N, Kimura M, Go E, Koyano F, Funayama M *et al.* (2012) PINK1 autophosphorylation upon membrane potential dissipation is essential for Parkin recruitment to damaged mitochondria. *Nat Commun* **3**, 1016.

- 59 Okatsu K, Uno M, Koyano F, Go E, Kimura M, Oka T, Tanaka K & Matsuda N (2013) A dimeric PINK1-containing complex on depolarized mitochondria stimulates Parkin recruitment. *J Biol Chem* **288**, 36372–36384.
- 60 Geisler S, Holmstrom KM, Skujat D, Fiesel FC, Rothfuss OC, Kahle PJ & Springer W (2010) PINK1/Parkin-mediated mitophagy is dependent on VDAC1 and p62/SQSTM1. *Nat Cell Biol* **12**, 119–131.
- 61 Narendra D, Kane LA, Hauser DN, Fearnley IM & Youle RJ (2010) p62/SQSTM1 is required for Parkin-induced mitochondrial clustering but not mitophagy; VDAC1 is dispensable for both. *Autophagy* **6**, 1090–1106.
- 62 Vives-Bauza C, Zhou C, Huang Y, Cui M, de Vries RL, Kim J, May J, Tocilescu MA, Liu W, Ko HS *et al.* (2010) PINK1-dependent recruitment of Parkin to mitochondria in mitophagy. *Proc Natl Acad Sci USA* **107**, 378–383.
- 63 Chaugule VK, Burchell L, Barber KR, Sidhu A, Leslie SJ, Shaw GS & Walden H (2011) Autoregulation of Parkin activity through its ubiquitin-like domain. *EMBO J* **30**, 2853–2867.
- 64 Kumar A, Aguirre JD, Condos TE, Martinez-Torres RJ, Chaugule VK, Toth R, Sundaramoorthy R, Mercier P, Knebel A, Spratt DE *et al.* (2015) Disruption of the autoinhibited state primes the E3 ligase parkin for activation and catalysis. *EMBO J* **34**, 2506–2521.
- 65 Shiba-Fukushima K, Imai Y, Yoshida S, Ishihama Y, Kanao T, Sato S & Hattori N (2012) PINK1-mediated phosphorylation of the Parkin ubiquitin-like domain primes mitochondrial translocation of Parkin and regulates mitophagy. *Sci Rep* **2**, 1002.
- 66 Kane LA, Lazarou M, Fogel AI, Li Y, Yamano K, Sarraf SA, Banerjee S & Youle RJ (2014) PINK1 phosphorylates ubiquitin to activate Parkin E3 ubiquitin ligase activity. *J Cell Biol* **205**, 143–153.
- 67 Kazlauskaitė A, Kondapalli C, Gurlay R, Campbell DG, Ritorto MS, Hofmann K, Alessi DR, Knebel A, Trost M & Muqit MM (2014) Parkin is activated by PINK1-dependent phosphorylation of ubiquitin at Ser65. *Biochem J* **460**, 127–139.
- 68 Koyano F, Okatsu K, Kosako H, Tamura Y, Go E, Kimura M, Kimura Y, Tsuchiya H, Yoshihara H, Hirokawa T *et al.* (2014) Ubiquitin is phosphorylated by PINK1 to activate parkin. *Nature* **510**, 162–166.
- 69 Okatsu K, Koyano F, Kimura M, Kosako H, Saeki Y, Tanaka K & Matsuda N (2015) Phosphorylated ubiquitin chain is the genuine Parkin receptor. *J Cell Biol* **209**, 111–128.
- 70 Ordureau A, Sarraf SA, Duda DM, Heo JM, Jedrychowski MP, Sviderskiy VO, Olszewski JL, Koerber JT, Xie T, Beausoleil SA *et al.* (2014) Quantitative proteomics reveal a feedforward mechanism for mitochondrial PARKIN translocation and ubiquitin chain synthesis. *Mol Cell* **56**, 360–375.
- 71 Shiba-Fukushima K, Inoshita T, Hattori N & Imai Y (2014) Lysine 63-linked polyubiquitination is dispensable for Parkin-mediated mitophagy. *J Biol Chem* **289**, 33131–33136.
- 72 Clague MJ & Urbe S (2017) Integration of cellular ubiquitin and membrane traffic systems: focus on deubiquitylases. *FEBS J* **284**, 1753–1766.
- 73 Yamano K, Matsuda N & Tanaka K (2016) The ubiquitin signal and autophagy: an orchestrated dance leading to mitochondrial degradation. *EMBO Rep* **17**, 300–316.
- 74 Lazarou M, Sliter DA, Kane LA, Sarraf SA, Wang C, Burman JL, Sideris DP, Fogel AI & Youle RJ (2015) The ubiquitin kinase PINK1 recruits autophagy receptors to induce mitophagy. *Nature* **524**, 309–314.
- 75 Heo JM, Ordureau A, Paulo JA, Rinehart J & Harper JW (2015) The PINK1-PARKIN mitochondrial ubiquitylation pathway drives a program of OPTN/NDP52 recruitment and TBK1 activation to promote mitophagy. *Mol Cell* **60**, 7–20.
- 76 Wei Y, Chiang WC, Sumpter R Jr, Mishra P & Levine B (2017) Prohibitin 2 is an inner mitochondrial membrane mitophagy receptor. *Cell* **168**, 224–238.e10.
- 77 Hamacher-Brady A, Brady NR, Logue SE, Sayen MR, Jinno M, Kirshenbaum LA, Gottlieb RA & Gustafsson AB (2007) Response to myocardial ischemia/reperfusion injury involves Bnip3 and autophagy. *Cell Death Differ* **14**, 146–157.
- 78 Chinnadurai G, Vijayalingam S & Gibson SB (2008) BNIP3 subfamily BH3-only proteins: mitochondrial stress sensors in normal and pathological functions. *Oncogene* **27** (Suppl 1), S114–S127.
- 79 Novak I, Kirkin V, McEwan DG, Zhang J, Wild P, Rozenknop A, Rogov V, Lohr F, Popovic D, Occhipinti A *et al.* (2010) Nix is a selective autophagy receptor for mitochondrial clearance. *EMBO Rep* **11**, 45–51.
- 80 Aerbajinai W, Giattina M, Lee YT, Raffeld M & Miller JL (2003) The proapoptotic factor Nix is coexpressed with Bcl-xL during terminal erythroid differentiation. *Blood* **102**, 712–717.
- 81 Melser S, Chatelain EH, Lavie J, Mahfouf W, Jose C, Obre E, Goorden S, Priault M, Elgersma Y, Rezvani HR *et al.* (2013) Rheb regulates mitophagy induced by mitochondrial energetic status. *Cell Metab* **17**, 719–730.
- 82 Chourasia AH, Tracy K, Frankenberger C, Boland ML, Sharifi MN, Drake LE, Sachleben JR, Asara JM, Locasale JW, Karczmar GS *et al.* (2015) Mitophagy defects arising from BNIP3 loss promote mammary tumor progression to metastasis. *EMBO Rep* **16**, 1145–1163.
- 83 Zhu Y, Massen S, Terenzio M, Lang V, Chen-Lindner S, Eils R, Novak I, Dikic I, Hamacher-Brady A &

- Brady NR (2013) Modulation of serines 17 and 24 in the LC3-interacting region of Bnip3 determines pro-survival mitophagy versus apoptosis. *J Biol Chem* **288**, 1099–1113.
- 84 Rogov VV, Suzuki H, Marinkovic M, Lang V, Kato R, Kawasaki M, Buljubasic M, Sprung M, Rogova N, Wakatsuki S *et al.* (2017) Phosphorylation of the mitochondrial autophagy receptor Nix enhances its interaction with LC3 proteins. *Sci Rep* **7**, 1131.
- 85 Liu L, Feng D, Chen G, Chen M, Zheng Q, Song P, Ma Q, Zhu C, Wang R, Qi W *et al.* (2012) Mitochondrial outer-membrane protein FUNDC1 mediates hypoxia-induced mitophagy in mammalian cells. *Nat Cell Biol* **14**, 177–185.
- 86 Chen G, Han Z, Feng D, Chen Y, Chen L, Wu H, Huang L, Zhou C, Cai X, Fu C *et al.* (2014) A regulatory signaling loop comprising the PGAM5 phosphatase and CK2 controls receptor-mediated mitophagy. *Mol Cell* **54**, 362–377.
- 87 Wu H, Xue D, Chen G, Han Z, Huang L, Zhu C, Wang X, Jin H, Wang J, Zhu Y *et al.* (2014) The BCL2L1 and PGAM5 axis defines hypoxia-induced receptor-mediated mitophagy. *Autophagy* **10**, 1712–1725.
- 88 Wu W, Tian W, Hu Z, Chen G, Huang L, Li W, Zhang X, Xue P, Zhou C, Liu L *et al.* (2014) ULK1 translocates to mitochondria and phosphorylates FUNDC1 to regulate mitophagy. *EMBO Rep* **15**, 566–575.
- 89 Chen Z, Liu L, Cheng Q, Li Y, Wu H, Zhang W, Wang Y, Sehgal SA, Siraj S, Wang X *et al.* (2017) Mitochondrial E3 ligase MARCH5 regulates FUNDC1 to fine-tune hypoxic mitophagy. *EMBO Rep* **18**, 495–509.
- 90 Kataoka T, Holler N, Micheau O, Martinon F, Tinel A, Hofmann K & Tschoep J (2001) Bcl-rambo, a novel Bcl-2 homologue that induces apoptosis via its unique C-terminal extension. *J Biol Chem* **276**, 19548–19554.
- 91 Edlich F & Lucke C (2011) From cell death to viral replication: the diverse functions of the membrane-associated FKBP38. *Curr Opin Pharmacol* **11**, 348–353.
- 92 Saita S, Shirane M & Nakayama KI (2013) Selective escape of proteins from the mitochondria during mitophagy. *Nat Commun* **4**, 1410.
- 93 Bhujabal Z, Birgisdottir AB, Sjøttem E, Brenne HB, Overvatn A, Habisov S, Kirkin V, Lamark T & Johansen T (2017) FKBP8 recruits LC3A to mediate Parkin-independent mitophagy. *EMBO Rep* **18**, 947–961.
- 94 Maguire JJ, Tyurina YY, Mohammadyani D, Kapralov AA, Anthonymuthu TS, Qu F, Amoscato AA, Sparvero LJ, Tyurin VA, Planas-Iglesias J *et al.* (2017) Known unknowns of cardiolipin signaling: the best is yet to come. *Biochim Biophys Acta* **1862**, 8–24.
- 95 Chu CT, Ji J, Dagda RK, Jiang JF, Tyurina YY, Kapralov AA, Tyurin VA, Yanamala N, Shrivastava IH, Mohammadyani D *et al.* (2013) Cardiolipin externalization to the outer mitochondrial membrane acts as an elimination signal for mitophagy in neuronal cells. *Nat Cell Biol* **15**, 1197–1205.
- 96 Kagan VE, Jiang J, Huang Z, Tyurina YY, Desbordes C, Cottet-Rousselle C, Dar HH, Verma M, Tyurin VA, Kapralov AA *et al.* (2016) NDPK-D (NM23-H4)-mediated externalization of cardiolipin enables elimination of depolarized mitochondria by mitophagy. *Cell Death Differ* **23**, 1140–1151.
- 97 Ban T, Ishihara T, Kohno H, Saita S, Ichimura A, Maenaka K, Oka T, Mihara K & Ishihara N (2017) Molecular basis of selective mitochondrial fusion by heterotypic action between OPA1 and cardiolipin. *Nat Cell Biol* **19**, 856–863.
- 98 Macdonald PJ, Stepanyants N, Mehrotra N, Mears JA, Qi X, Sesaki H & Ramachandran R (2014) A dimeric equilibrium intermediate nucleates Drp1 reassembly on mitochondrial membranes for fission. *Mol Biol Cell* **25**, 1905–1915.
- 99 Montessuit S, Somasekharan SP, Terrones O, Lucken-Ardjomande S, Herzig S, Schwarzenbacher R, Manstein DJ, Bossy-Wetzel E, Basanez G, Meda P *et al.* (2010) Membrane remodeling induced by the dynamin-related protein Drp1 stimulates Bax oligomerization. *Cell* **142**, 889–901.
- 100 Ivatt RM, Sanchez-Martinez A, Godena VK, Brown S, Ziviani E & Whitworth AJ (2014) Genome-wide RNAi screen identifies the Parkinson disease GWAS risk locus SREBF1 as a regulator of mitophagy. *Proc Natl Acad Sci USA* **111**, 8494–8499.
- 101 Ding WX, Ni HM, Li M, Liao Y, Chen X, Stolz DB, Dorn GW 2nd & Yin XM (2010) Nix is critical to two distinct phases of mitophagy, reactive oxygen species-mediated autophagy induction and Parkin-ubiquitin-p62-mediated mitochondrial priming. *J Biol Chem* **285**, 27879–27890.
- 102 Gao F, Chen D, Si J, Hu Q, Qin Z, Fang M & Wang G (2015) The mitochondrial protein BNIP3L is the substrate of PARK2 and mediates mitophagy in PINK1/PARK2 pathway. *Hum Mol Genet* **24**, 2528–2538.
- 103 Zhang T, Xue L, Li L, Tang C, Wan Z, Wang R, Tan J, Tan Y, Han H, Tian R *et al.* (2016) BNIP3 protein suppresses PINK1 kinase proteolytic cleavage to promote mitophagy. *J Biol Chem* **291**, 21616–21629.
- 104 Koentjoro B, Park JS & Sue CM (2017) Nix restores mitophagy and mitochondrial function to protect against PINK1/Parkin-related Parkinson's disease. *Sci Rep* **7**, 44373.
- 105 Wong PM, Puente C, Ganley IG & Jiang X (2013) The ULK1 complex: sensing nutrient signals for autophagy activation. *Autophagy* **9**, 124–137.

- 106 Papinski D & Kraft C (2016) Regulation of autophagy by signaling through the Atg1/ULK1 complex. *J Mol Biol* **428**, 1725–1741.
- 107 Hurley JH & Young LN (2017) Mechanisms of autophagy initiation. *Annu Rev Biochem* **86**, 225–244.
- 108 Kundu M, Lindsten T, Yang CY, Wu J, Zhao F, Zhang J, Selak MA, Ney PA & Thompson CB (2008) Ulk1 plays a critical role in the autophagic clearance of mitochondria and ribosomes during reticulocyte maturation. *Blood* **112**, 1493–1502.
- 109 Honda S, Arakawa S, Nishida Y, Yamaguchi H, Ishii E & Shimizu S (2014) Ulk1-mediated Atg5-independent macroautophagy mediates elimination of mitochondria from embryonic reticulocytes. *Nat Commun* **5**, 4004.
- 110 Itakura E, Kishi-Itakura C, Koyama-Honda I & Mizushima N (2012) Structures containing Atg9A and the ULK1 complex independently target depolarized mitochondria at initial stages of Parkin-mediated mitophagy. *J Cell Sci* **125**, 1488–1499.
- 111 Gilkerson RW, De Vries RL, Lebot P, Wikstrom JD, Torgyekes E, Shiriha OS, Przedborski S & Schon EA (2012) Mitochondrial autophagy in cells with mtDNA mutations results from synergistic loss of transmembrane potential and mTORC1 inhibition. *Hum Mol Genet* **21**, 978–990.
- 112 Bach M, Larance M, James DE & Ramm G (2011) The serine/threonine kinase ULK1 is a target of multiple phosphorylation events. *Biochem J* **440**, 283–291.
- 113 Egan DF, Shackelford DB, Mihaylova MM, Gelino S, Kohnz RA, Mair W, Vasquez DS, Joshi A, Gwinn DM, Taylor R *et al.* (2011) Phosphorylation of ULK1 (hATG1) by AMP-activated protein kinase connects energy sensing to mitophagy. *Science* **331**, 456–461.
- 114 Mack HI, Zheng B, Asara JM & Thomas SM (2012) AMPK-dependent phosphorylation of ULK1 regulates ATG9 localization. *Autophagy* **8**, 1197–1214.
- 115 Shang L, Chen S, Du F, Li S, Zhao L & Wang X (2011) Nutrient starvation elicits an acute autophagic response mediated by Ulk1 dephosphorylation and its subsequent dissociation from AMPK. *Proc Natl Acad Sci USA* **108**, 4788–4793.
- 116 Kim J, Kundu M, Viollet B & Guan KL (2011) AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol* **13**, 132–141.
- 117 Zhu H, Foretz M, Xie Z, Zhang M, Zhu Z, Xing J, Leclerc J, Gaudry M, Viollet B & Zou MH (2014) PRKAA1/AMPKalpha1 is required for autophagy-dependent mitochondrial clearance during erythrocyte maturation. *Autophagy* **10**, 1522–1534.
- 118 Tian W, Li W, Chen Y, Yan Z, Huang X, Zhuang H, Zhong W, Chen Y, Wu W, Lin C *et al.* (2015) Phosphorylation of ULK1 by AMPK regulates translocation of ULK1 to mitochondria and mitophagy. *FEBS Lett* **589**, 1847–1854.
- 119 Nguyen TN, Padman BS, Usher J, Oorschot V, Ramm G & Lazarou M (2016) Atg8 family LC3/GABARAP proteins are crucial for autophagosome-lysosome fusion but not autophagosome formation during PINK1/Parkin mitophagy and starvation. *J Cell Biol* **215**, 857–874.
- 120 Tsuboyama K, Koyama-Honda I, Sakamaki Y, Koike M, Morishita H & Mizushima N (2016) The ATG conjugation systems are important for degradation of the inner autophagosomal membrane. *Science* **354**, 1036–1041.
- 121 McWilliams TG, Prescott AR, Allen GF, Tamjar J, Munson MJ, Thomson C, Muqit MM & Ganley IG (2016) mito-QC illuminates mitophagy and mitochondrial architecture *in vivo*. *J Cell Biol* **214**, 333–345.
- 122 Sun N, Yun J, Liu J, Malide D, Liu C, Rovira II, Holmstrom KM, Fergusson MM, Yoo YH, Combs CA *et al.* (2015) Measuring *in vivo* mitophagy. *Mol Cell* **60**, 685–696.
- 123 Gottlieb RA & Stotland A (2015) MitoTimer: a novel protein for monitoring mitochondrial turnover in the heart. *J Mol Med (Berl)* **93**, 271–278.
- 124 Kuma A, Komatsu M & Mizushima N (2017) Autophagy-monitoring and autophagy-deficient mice. *Autophagy* **13**, 1619–1628.
- 125 McWilliams TG & Ganley IG (2016) Life in lights: tracking mitochondrial delivery to lysosomes *in vivo*. *Autophagy* **12**, 2506–2507.
- 126 Pickrell AM, Huang CH, Kennedy SR, Ordureau A, Sideris DP, Hoekstra JG, Harper JW & Youle RJ (2015) Endogenous parkin preserves dopaminergic substantia nigral neurons following mitochondrial DNA mutagenic stress. *Neuron* **87**, 371–381.
- 127 Pagliarini DJ, Calvo SE, Chang B, Sheth SA, Vafai SB, Ong SE, Walford GA, Sugiana C, Boneh A, Chen WK *et al.* (2008) A mitochondrial protein compendium elucidates complex I disease biology. *Cell* **134**, 112–123.
- 128 Ishihara M, Urushido M, Hamada K, Matsumoto T, Shimamura Y, Ogata K, Inoue K, Taniguchi Y, Horino T, Fujieda M *et al.* (2013) Sestrin-2 and BNIP3 regulate autophagy and mitophagy in renal tubular cells in acute kidney injury. *Am J Physiol Renal Physiol* **305**, F495–F509.
- 129 Jiang M, Wei Q, Dong G, Komatsu M, Su Y & Dong Z (2012) Autophagy in proximal tubules protects against acute kidney injury. *Kidney Int* **82**, 1271–1283.
- 130 Clark SL Jr (1957) Cellular differentiation in the kidneys of newborn mice studies with the electron microscope. *J Biophys Biochem Cytol* **3**, 349–362.
- 131 Hansell P, Welch WJ, Blantz RC & Palm F (2013) Determinants of kidney oxygen consumption and their relationship to tissue oxygen tension in diabetes and hypertension. *Clin Exp Pharmacol Physiol* **40**, 123–137.

- 132 Hall AM, Unwin RJ, Parker N & Duchon MR (2009) Multiphoton imaging reveals differences in mitochondrial function between nephron segments. *J Am Soc Nephrol* **20**, 1293–1302.
- 133 Mizushima N, Yamamoto A, Matsui M, Yoshimori T & Ohsumi Y (2004) *In vivo* analysis of autophagy in response to nutrient starvation using transgenic mice expressing a fluorescent autophagosome marker. *Mol Biol Cell* **15**, 1101–1111.
- 134 Davis CH, Kim KY, Bushong EA, Mills EA, Boassa D, Shih T, Kinebuchi M, Phan S, Zhou Y, Bihlmeyer NA *et al.* (2014) Transcellular degradation of axonal mitochondria. *Proc Natl Acad Sci USA* **111**, 9633–9638.